

Understanding Transcriptional Regulatory Redundancy by Learnable Global Subset Perturbations

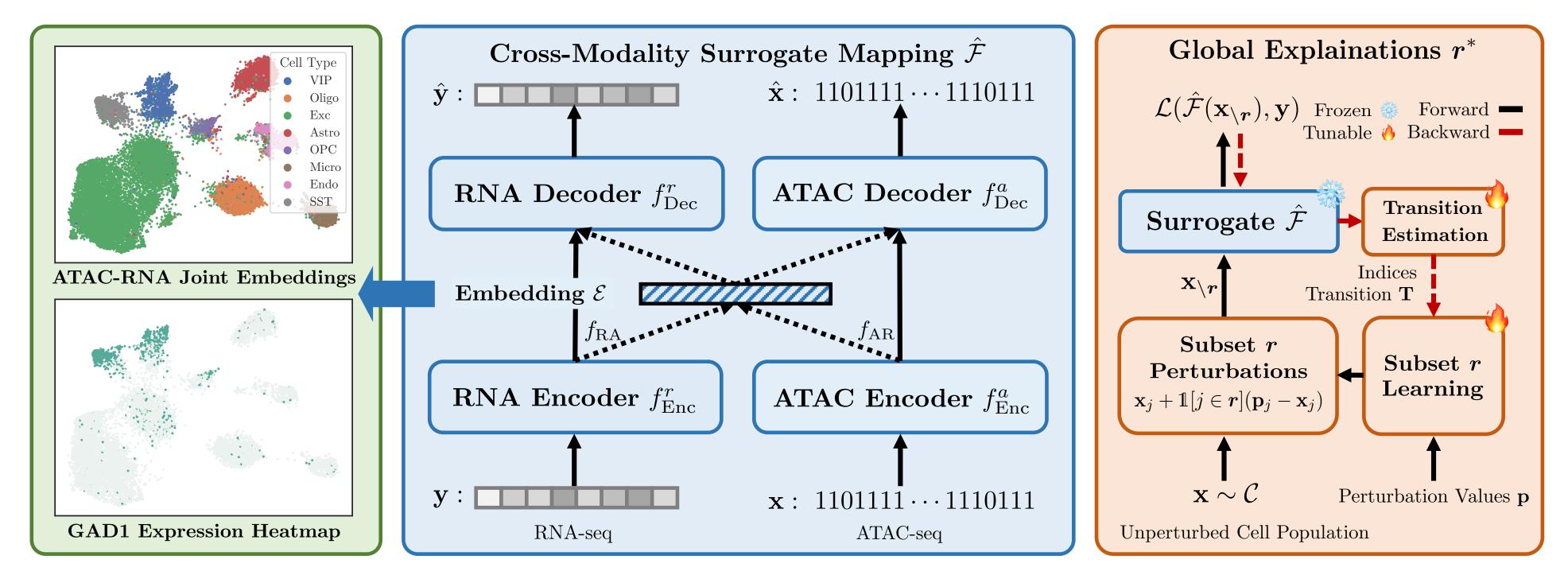
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Introduction to GRIDS

Transcriptional regulation through cis-regulatory elements (CREs) is crucial for numerous biological functions, and its disruption can potentially lead to various diseases. It is well known that these CREs often exhibit redundancy, enabling them to compensate for one another in response to external disturbances. This underscores the need for methods to identify CRE sets that collaboratively regulate gene expression effectively. To address this:

We present GRIDS, a computational method framing



- CRE dissection as a global feature explanation task.
- GRIDS first builds a differentiable surrogate function to approximate gene regulation and enable single-cell modality translation.
- It then uses learnable perturbations in a state transition framework to provide global explanations, efficiently exploring the feature landscape.

Preliminary of Single-Cell Data

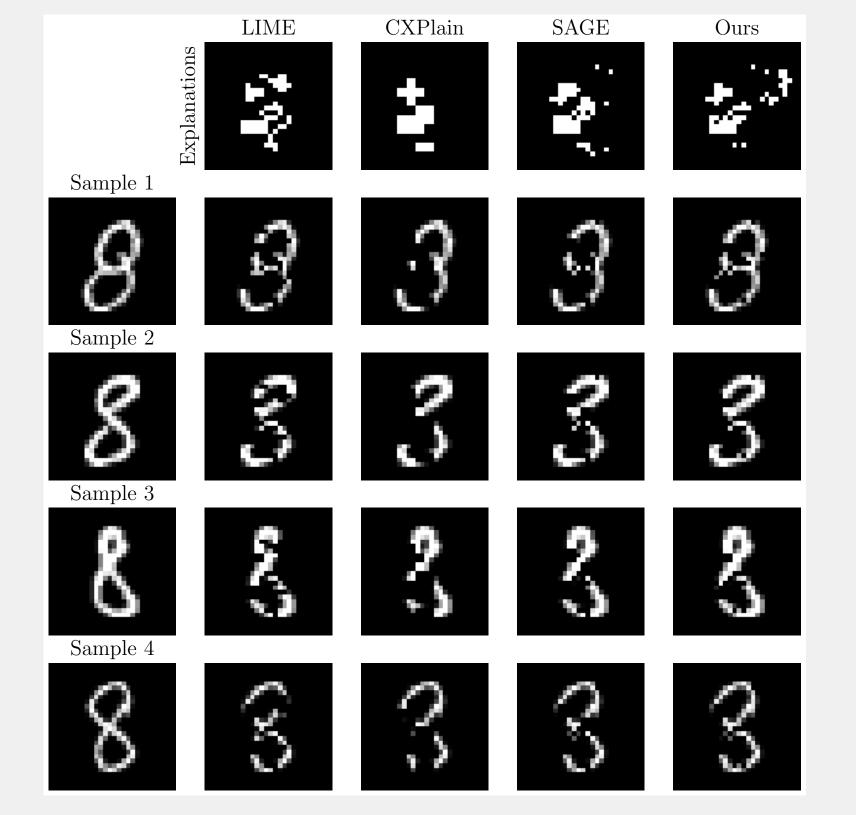
The CRE is represented by the ATAC-seq binary vector $\mathbf{x} \in \{0, 1\}^{d_a}$, where each dimension indicates a chromosome peak's state ("1" for open, "0" for closed). Typically, $d_a > 10^5$. Gene expression (RNA-seq) regulated by the CRE is denoted as $\mathbf{y} \in \mathbb{R}^{d_r}$, where d_a and d_r represent the number of peaks and genes, respectively. A single-cell multi-omics dataset consists of N cells $\mathcal{C} =$ $\{c^{(1)}, c^{(2)}, \dots, c^{(N)}\}$, with each cell $c^{(i)} = (\mathbf{x}^{(i)}, \mathbf{y}^{(i)})$ containing an ATAC-seq vector and its corresponding RNAseq vector. Each cell also has a label $\ell^{(i)} \in \{1, \ldots, T\}$ indicating its type among T classes.

Regulatory Redundancy Problem

Figure 1. Overview of our proposed GRIDS method. It comprises two steps: training a cross-modality surrogate model and using a global explanation method to dissect regulatory redundancy.

Demo on MNIST

To evaluate GRIDS's global feature importance estimation, we tested its ability to identify key features in MNIST images. A binary classification model was trained to distinguish digits 8 and 3, achieving 97.9% accuracy on the test set. Various explanation methods were then used to identify the top L = 64 important pixels, masking them to zero (p = 0). All methods produced subsets of L pixels based on their importance scores.



Experiment on Brain Data (Continued)

Type L	Method	Avg. Δ	Rel. ∆ (%)
	Random	-0.448	-0.009
	Saliency	-18.822	-0.915
VIP-100 10) SmoothGrad	-18.424	-0.927
	FIMAP	-56.469	-3.087
	GRIDS	-64.016	-3.827
	Random	-0.333	-0.008
	Saliency	-42.372	-1.941
Microglia-100 10	SmoothGrad	-44.125	-2.073
	FIMAP	-115.092	-5.863
	GRIDS	-141.339	-7.466

Table 2. Cell-type-focused benchmark results in VIP and Microglia by comparing expression degradation of highly expressed genes after masking CRE features in the global explanation subset r.

Gene expression is regulated by CREs through complex biological processes, modeled as $y = \mathcal{F}(x)$, where $\mathcal{F}(\mathbf{x}): \mathbb{R}^{d_a} \to \mathbb{R}^{d_r}$. Due to high experimental costs, frequent queries of the black-box function \mathcal{F} are challenging. Regulatory redundancy dissection seeks a subset of L peak indices $r = \{r_1, \ldots, r_L\}$ within the CRE (i.e., features in ATAC-seq $\mathbf{x}_r \equiv {\mathbf{x}_i | j \in \mathbf{r}}$ that are critical for regulating gene expression across a cell population.

Global Feature Explanations for Regulatory Redundancy Dissection

We propose an in silico computational method by modeling it within a global feature explanation framework. Conventionally, global explanation is defined by how much a model's performance degrades over an observed population of samples when features are removed. In the context of regulatory redundancy, the global explanation objective can be expressed as

$m{r}^* = \operatorname*{argmin}_{m{r}} \mathbb{E}_{\mathbf{c} \sim \mathcal{C}}[\mathcal{L}(\mathcal{F}(\mathbf{x}_{\setminus m{r}}), \mathbf{y})]$

where \mathcal{L} is a loss measurement for expected gene expression degradation. $\mathbf{x}_{\setminus r}$ denotes the perturbed CREs induced by r, replacing the original feature x_r with preset perturbation values $\mathbf{p} \in \mathbb{R}^{d_a}$.

Figure 2. An examination of the most significant L = 64 pixels identified by various methods. Our subset perturbation learning method can find a similar combinatorial pattern as SAGE.

Experiment on Brain Data

We curated a set of deeply-sequenced single-cell multimodal data from postmortem human. We then evaluated the performance of GRIDS to dissect multi-CRE-togene regulatory redundancy by generating global feature importance explanations in the high-throughput single-cell multi-omics data. The global explanations were learned in the training set and then evaluated its performance on the test set. SmoothGrad FIMAP GRIDS Random Saliency **Type** Avg. \triangle Rel. $\triangle(\%)$ Avg. \triangle Rel. $\triangle(\%)$ Avg. \triangle Rel. $\triangle(\%)$ Avg. \triangle Rel. $\triangle(\%)$ Avg. \triangle Rel. $\triangle(\%)$ -2.155-0.601-13.502-5.837-2.163-0.621

Method	$L = 10$ HR \uparrow SHR \uparrow		$L = 128$ HR \uparrow SHR \uparrow	
Saliency	0.00	0.00	6.25	18.75
SmoothGrad	0.00	0.00	0.00	18.75
FIMAP	12.50	12.50	18.75	56.25
GRIDS	18.75	25.00	31.25	68.75

Table 3. The hit ratio of direct CRE-to-gene interactions.

Highlights

- We extend feature explanation techniques to scientific discovery on single-cell data.
- Through comprehensive benchmarking, GRIDS demonstrates superior explanatory capabilities compared to other leading methods.
- Moreover, GRIDS's global explanations reveal intricate regulatory redundancies across cell types and states, underscoring its potential to advance our understanding of cellular regulation in biological research.

Subset Transition Matrix for Gradient Estimation

- gradient w.r.t the input embedding $\mathbf{G} = \partial \mathbb{E}_{\mathbf{c} \sim \mathcal{C}} [\mathcal{L}(\hat{\mathcal{F}}(\mathbf{x}_{\backslash \mathbf{r}}), \mathbf{y})] / \partial \mathbf{W}^{a}_{\mathrm{Emb}}(\mathbf{x}_{\backslash \mathbf{r}})$
- construct the transition matrix $\mathbf{T} \in \mathbb{R}^{L \times d_a}$ via first-order approximation
- $T_{i,j}$ represents the advantage of replacing index r_i with j

 $\mathbf{d}_{j} = \mathbf{G}_{j} \cdot (\mathbf{W}^{a}_{\mathrm{Emb}}(\mathbf{p})_{j} - \mathbf{W}^{a}_{\mathrm{Emb}}(\mathbf{x})_{j})$ $\mathbf{T}_{i,j} = \mathbf{1}[j \notin \mathbf{r}]\mathbf{d}_j - \mathbf{1}[j \neq r_i]\mathbf{d}_{r_i}$

Endo	-1.073	-0.138 -4.974	-0.372 -9.726	-0.995	-38.997	-9.303 -57.477 -11.816
Micro	-0.012	-0.026 -23.757	-1.545 -32.944	-2.083	-73.752	-6.248 -90.607 -7.671
OPC	+0.823	-0.087 -54.645	-2.338 - 48.438	-2.067	-77.167	-6.260 -96.661 -8.256
Oligo	-0.058	+0.026 -0.558	-0.173 -0.939	-0.220	-10.917	-4.252 -16.760 -6.896
SST	+0.159	+0.080 -5.201	-2.006 -5.201	-2.006	-16.453	-5.660 -17.677 -6.365
VIP	+0.012	+0.001 -0.654	-1.189 -0.634	-1.160	-2.732	-3.797 -6.804 -7.195
Avg.	+0.016	-0.021 -12.988	-1.209 - 13.519	-1.290	-30.268	-5.367 - 39.103 - 7.300
Astro	-1.793	-0.533 -15.511	-4.853 - 18.505	-6.217	-82.565	-24.766 -100.556 -34.633
Endo	+2.554	+0.468 - 46.160	-6.217 -52.383	-7.893	-252.338	-41.790 -259.920 -44.601
Micro	-9.091	-0.490 - 131.512	-9.122 - 145.561	-10.116	-451.210	-39.695 - 470.430 - 44.114
OPC	-1.848	-0.165 - 193.739	-10.260 - 186.235	-9.891 -	-415.231	-35.687 -392.326 -36.380
Oligo	-1.134	-0.211 - 19.809	-6.382 -21.136	-7.630	-69.460	-28.175 -93.518 -38.982
SST	-1.681	-0.615 -33.589	-11.675 -32.275	-11.115	-86.191	-29.198 - 93.772 - 33.708
VIP	+0.071	+0.002 -4.014	-4.876 -3.872	-4.782	-13.054	-16.757 -19.703 -27.221
Avg.	-1.843	-0.237 -68.620	-7.618 -70.292	-8.212	-202.368	-30.787 - 209.583 - 36.893

Table 1. Gene-focused benchmark results by comparing expression drops of marker genes across all cell types (upper: L = 10, bottom: L = 128).





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